# A Novel Countermeasure Against Ionizing Radiation-Induced Bone Loss

Fei Wei<sup>1</sup>, Craig J. Neal<sup>2</sup>, Tamil Selvan Sakthivel<sup>2</sup>, Mahmoud Omer<sup>1</sup>, Amitava Adhikary<sup>3</sup>, Samuel Ward<sup>3</sup>, Khoa Minh Ta<sup>4</sup>, Samuel Moxon<sup>4</sup>, Marco Molinari<sup>4</sup>, Vee San Cheong<sup>5</sup>, Jackson Asiatico<sup>6</sup>, Michael Kinzel<sup>6</sup> Sergey N. Yarmolenko<sup>7</sup>, Nina Orlovskaya<sup>6</sup>, Ranajay Ghosh<sup>6</sup>, Sudipta Seal<sup>2</sup>, Melanie Coathup<sup>1</sup>





# **Background – Exposure of Bone to Ionizing Radiation**

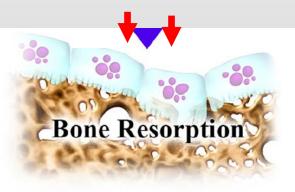
High Ca content  $\rightarrow$  30 - 40% more ionizing radiation (IR) absorption in bone<sup>1</sup>

#### **X** IR-induced bone toxicity:

- A Bone Pain
- Bone Atrophy
- BMD
- Osteoporosis
- Fracture incidence<sup>2</sup>
- Bone repair delayed & incomplete

Major health concern with no effective prophylactic

Macrophage (M1)
 Collagen dysregulation
 Microvessel necrosis
 Vascular thrombosis
 Osteoblast downregulation
 Osteoclast upregulation





<sup>1</sup>Curi et al. J Oral Max Surg, 2016, <sup>2</sup>Yaprak et al. BMC Cancer, 2018, <sup>3</sup>Baxter et al. 2005 Nov 23; 294(20):2587-93

## **Reactive Oxygen Species (ROS)**

In presence of oxygen many radicals form during IR-exposure e.g., OH<sup>•</sup>, H<sup>•</sup>, H<sub>2</sub>, H<sup>+</sup>, H<sub>2</sub>O<sub>2</sub>, O<sub>2</sub><sup>•-</sup>/HO<sub>2</sub><sup>•</sup>, organic radicals

 $\checkmark$  OH• hydroxyl radical  $\rightarrow$  most damaging.  $\frac{2}{3}$  DNA damage to cells, reacts with almost every organic biomolecule including DNA

 $\sqrt{O_2^{-}}$  easily generated, major culprit, precursor to most other harmful ROS

 $\checkmark$  H<sub>2</sub>O<sub>2</sub> – more toxic than O<sub>2</sub><sup>•-</sup>, directly generates OH<sup>•</sup> (Fenton reaction)

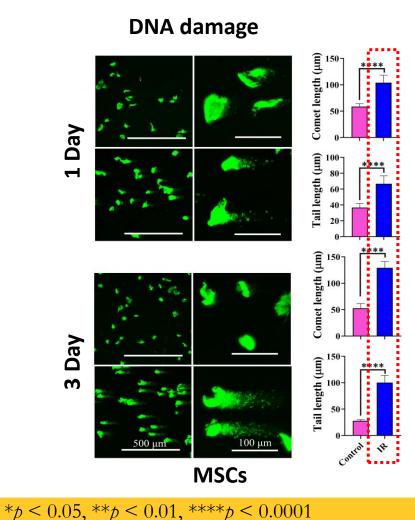
ROS micro-distribute known as "bystander effect"

- X Protein carbonylation
- X Lipid peroxidation
- X Spontaneous gene mutations
- X Neoplastic transformation

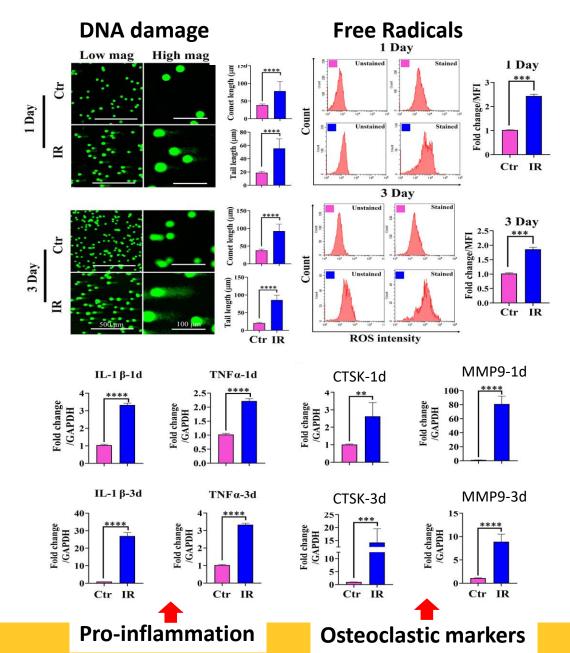


#### Ionizing radiation damages hBMSCs and macrophages

hBMSCs

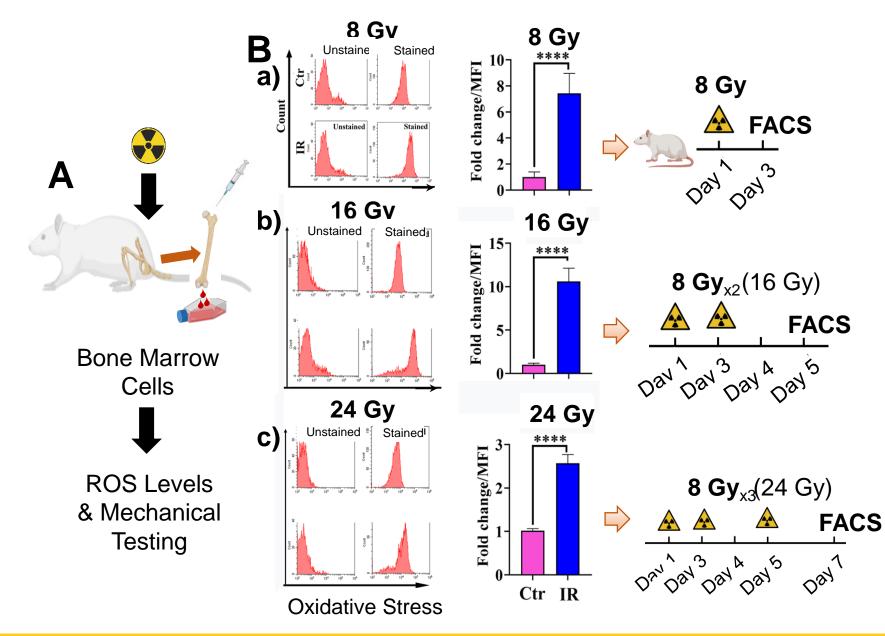


Macrophages

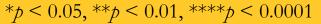




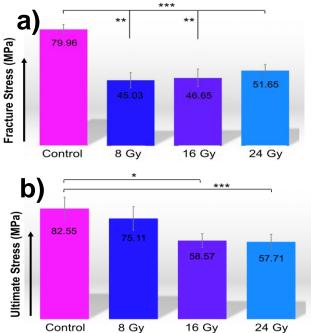
#### **Effect of Incremental IR Doses on Bone**

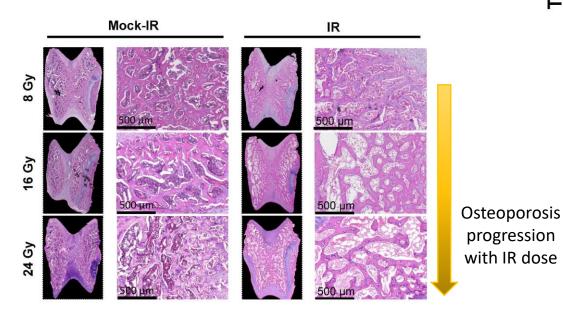


Ionizing radiation causes immediate ROS formation







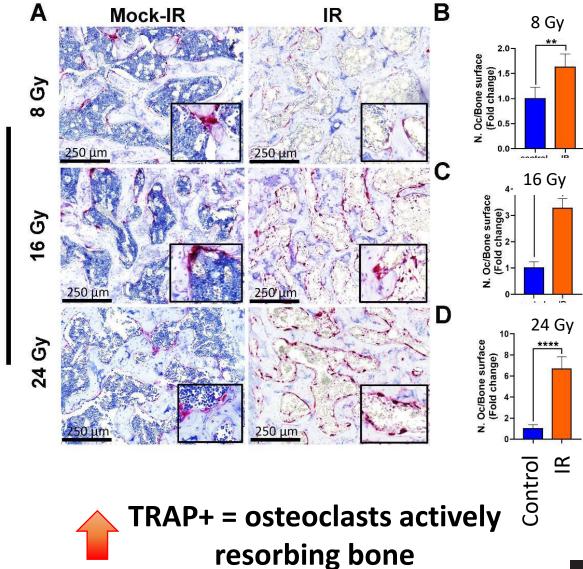


✓ Significant reduction in bone strength after first dose.



progression

with IR dose

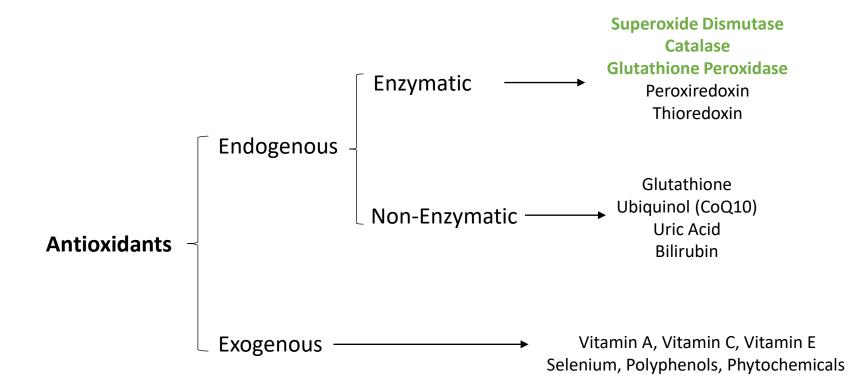




Healthy Bone

## **Antioxidant Enzymes**

 Cells counteract oxidative stress via radical scavenging by endogenous antioxidant systems *in situ* or by exogenous sources supplied through our diet

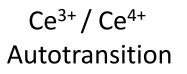


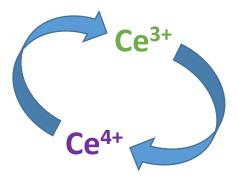
System becomes overwhelmed during oxidative stress
 Antioxidant biomaterials able to scavenge harmful free radicals and restore a healthy cellular redox balance are of growing interest



# **Cerium Oxide Nanoparticles (CeONPs)**

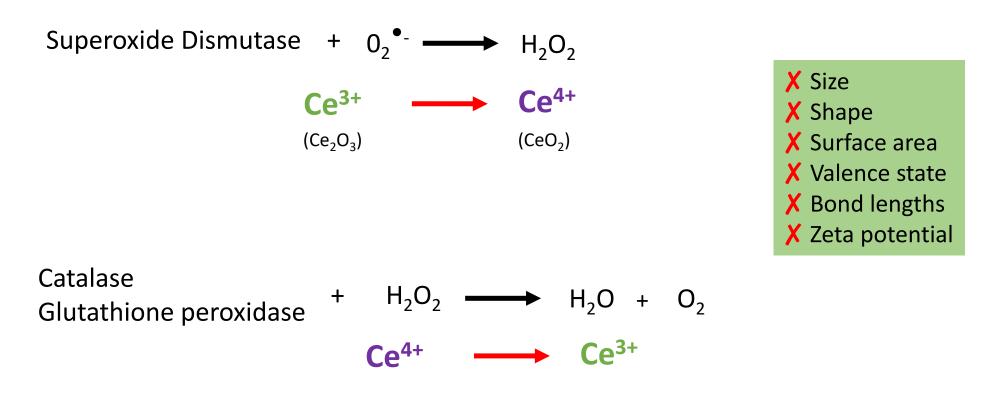
- Rare earth metals are a promising strategic resource
- 4f orbitals imparts unique catalytic, magnetic and electronic properties that are not possible with transition and main group metals
- Cerium oxide nanoparticles (CeONPs) new generation of Nanozyme – "artificial enzyme"
- Mimics multiple endogenous antioxidants able to scavenge almost all types of noxious reactive species
- Confirmed to outperform endogenous antioxidants







#### Cerium Oxide Nanoparticles (CeONPs) mimic activity of *multiple* endogenous antioxidant enzymes

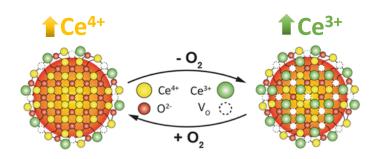


**Free radical scavenging** 



**Q1:** Will a nanozyme designed to target  $H_2O_2$  relative to  $O_2^{\bullet-}$  further increase the radioprotective effect of CeONPs to cells *in vitro*?

**Q2:** When administered into a rat model *in vivo*, is IR-induced DNA damage and subsequent bone loss prevented?



# Study Aim

To investigate the radioprotective effectiveness of two nanozymes: designed for greater relative (i) CAT (Ce<sup>4+</sup>) or (ii) SOD (Ce<sup>3+</sup>) activity following irradiation-induced damage *in vitro* and *in vivo*.



### **Study Hypotheses**

H1: Pre-treatment of cells with CeONPs prior to IR, will protect primary human bone marrow derived stem cells (hBMSCs) and RAW 264.7 macrophages by targeted scavenging of  $H_2O_2$  (and OH<sup>•</sup>) and will:

- $\circ~$  Reduce DNA damage and senescence.
- Increase proliferation, osteogenic differentiation and bone mineral deposition of hBMSCs.
- Reduce inflammatory and osteoclastic marker expression in macrophages.

**H2:** Following IR-induced damage and when administered to rats, bone will maintain its volume, architecture and strength.



## **Methods** - CeONP synthesis & characterization

#### **CeONP Synthesis**

(1) Wet Chemical technique<sup>6</sup>: lower fraction of Ce<sup>4+</sup> surface sites relative to Ce<sup>3+</sup>
 (2) Forced Hydrolysis<sup>7</sup>: higher fraction of Ce<sup>4+</sup> surface sites relative to Ce<sup>3+</sup>

#### **CeONP** characterization

- High-resolution transmission electron microscopy (HRTEM) (particle size)
- Dynamic Light Scattering (DLS) (hydrodynamic radius)
- o Zeta sizer (surface charge)
- X-Ray photoelectron spectroscopy (XPS) (quantify Ce<sup>3+</sup> & Ce<sup>4+</sup> fractions)
- $\circ~$  SOD (O2 -) and Catalase (H2O2) assays
- Electron paramagnetic resonance (specificity of Ce<sup>3+</sup> & Ce<sup>4+</sup> to O<sub>2</sub><sup>•-</sup>)
- Density Functional Theory (specificity of Ce<sup>3+</sup> & Ce<sup>4+</sup> to H<sub>2</sub>O<sub>2</sub>)

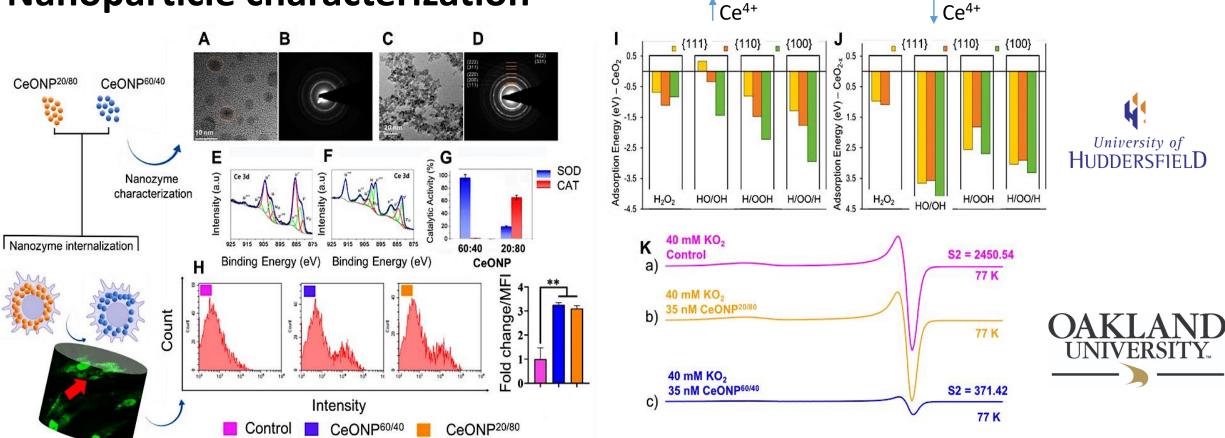
#### **CeONP Cellular Uptake**

- FITC-labelled CeONPs: imaged using confocal microscopy at 24h
- Quantified cellular internalization using flow cytometry





## Nanoparticle characterization



- ✓ EPR: Ce<sup>3+</sup> surface sites selectively neutralize  $O_2^{\bullet-}$
- $\checkmark \uparrow$  Ce<sup>3+</sup> increases ROS interaction with surfaces
- $\checkmark$   $\land$  Ce<sup>3+</sup> has greater scavenging activity than Ce<sup>4+</sup>
- ✓ OH• (HO/OH) only form on CeO<sub>2</sub> surfaces but are scavenged on CeO<sub>2-x</sub> surfaces Through Ce<sup>3+</sup> → Ce<sup>4+</sup>

UCF



#### (↑Ce<sup>3+</sup>) (↑Ce<sup>4+</sup>) In vitro Methods — in vitro analyses of CeONP<sup>60/40</sup> and CeONP<sup>20/80</sup>

- Material characterization (HRTEM, XPS, etc)
- DNA damage to hBMSCs and RAW 264.7 macrophages (7 Gy)
  - ✓ Alkaline Comet Assay<sup>®</sup>, 3d post-IR
- Intracellular ROS generation in hBMSCs (7 Gy)

✓ Cellular ROS Assay Kit counter-stained with MitoSpy<sup>®</sup>, 24h post-IR

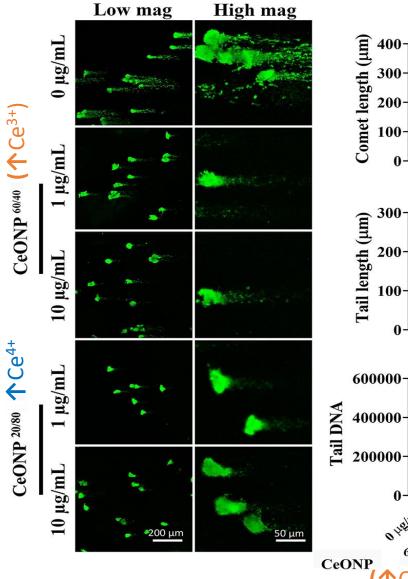
• Intracellular  $O_2^{\bullet-}$  levels (7 Gy)

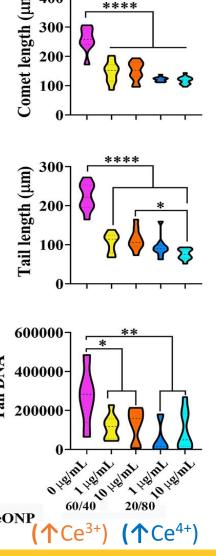
✓ MitoSOX<sup>®</sup> Red mitochondrial superoxide indicator kit

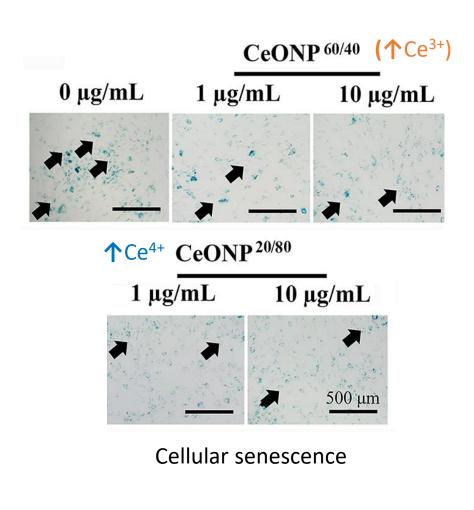
- Gene expression (qRT-PCR), and protein release (ELISA) (7 Gy)
  - ✓ CAT, SOD and GPX in hBMSCs, 24h post-IR
  - $\checkmark$  Pro-inflammatory cytokine expression (IL-1 $\beta$  and IL-6) in macrophages
  - ✓ Bone-resorbing osteoclastic differentiation markers (RANKL and CTSK) in macrophages
- IR-induced cellular senescence (7 Gy)
  - $\checkmark$   $\beta$ -galactosidase ( $\beta$ -gal) staining kit, hBMSCs, 28-days post-IR
- O Bone forming osteogenic differentiation in hBMSCs and bone mineral deposition (7 Gy)
  ✓ Alizarin Red assay, 28 days post-IR



#### CeONP<sup>60/40</sup> & CeONP<sup>20/80</sup> reduces IR-induced DNA damage and cell senescence in hBMSCs

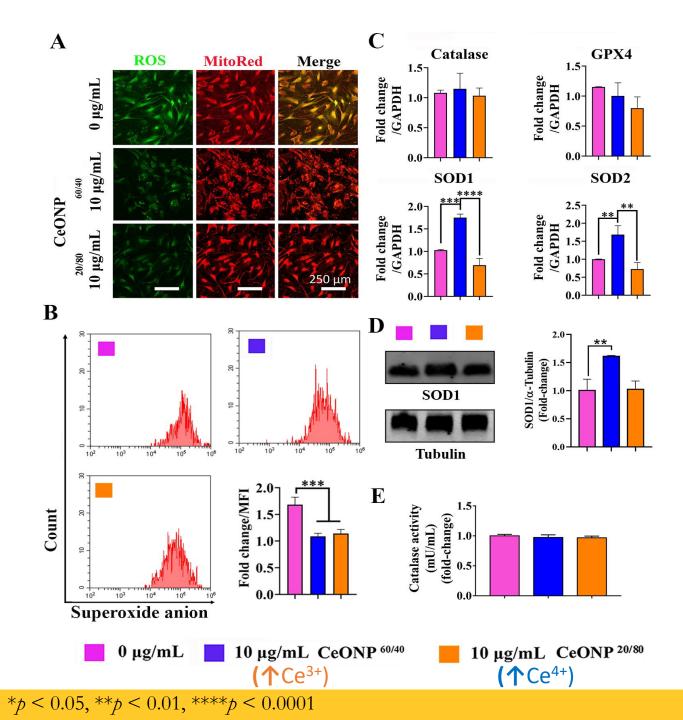








\*p < 0.05, \*\*p < 0.01, \*\*\*\*p < 0.0001

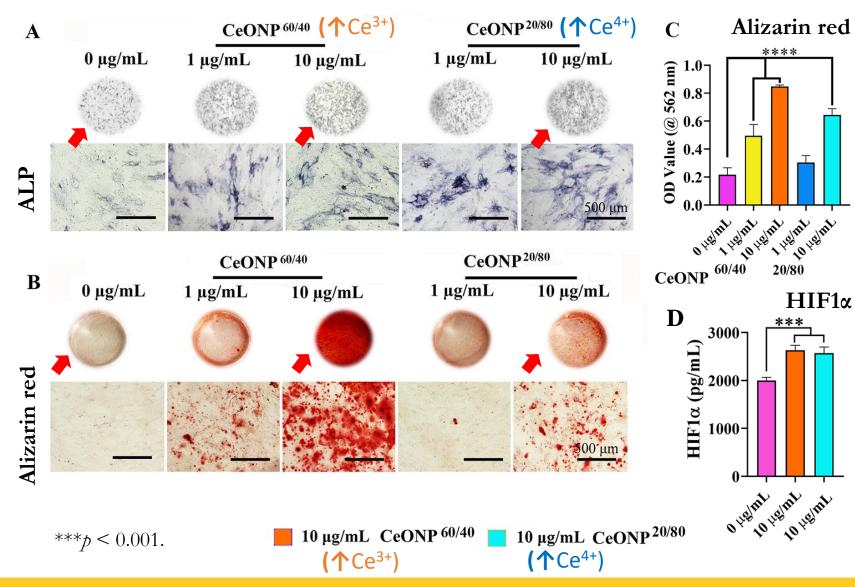


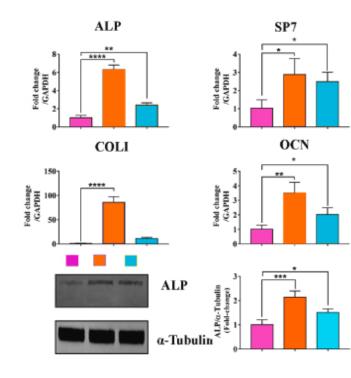
CeONP<sup>60/40</sup> and CeONP<sup>20/80</sup> reduce radiation-induced intracellular ROS/superoxide anion generation

 ✓ ↑Ce<sup>3+</sup> increases SOD but not Catalase or GPX gene expression in hBMSCs
 ✓ Both formulations scavenge O<sub>2</sub><sup>•−</sup> to a similar degree



# Both CeONP<sup>60/40</sup> and CeONP<sup>20/80</sup> liberates osteoblastogenesis following irradiation, but **^**Ce<sup>3+</sup> promotes a greater response

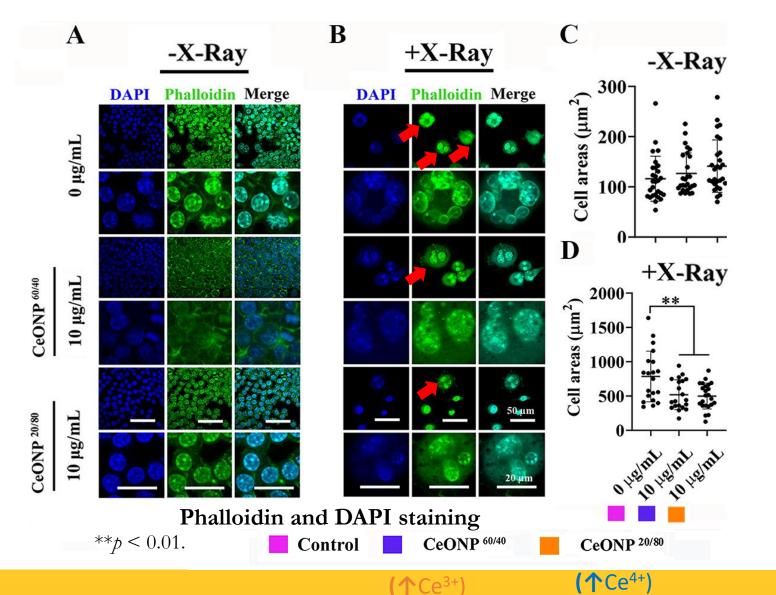




Ce<sup>3+</sup> upregulates osteogenic protein gene expression

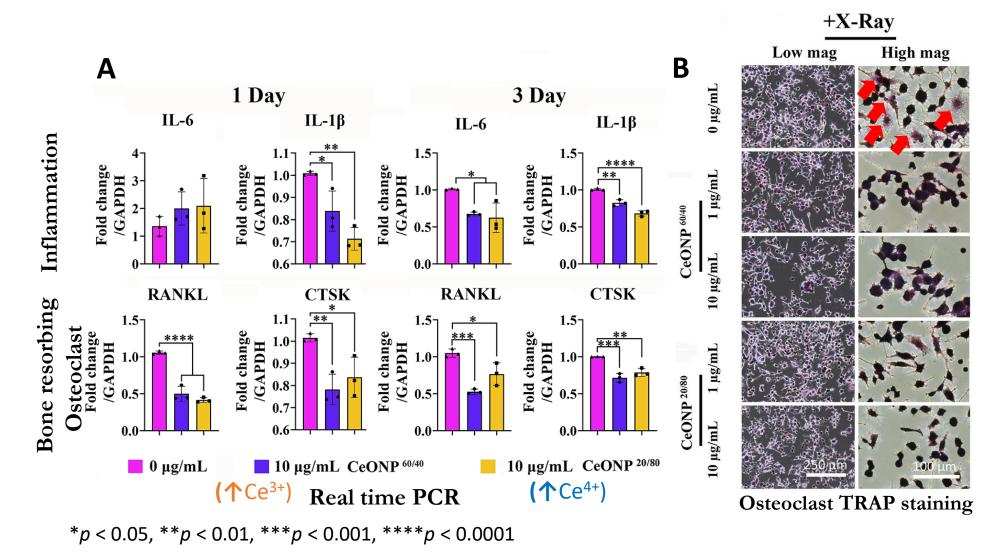


#### CeONP<sup>60/40</sup> and CeONP<sup>20/80</sup> repress osteoclast-like giant cell formation following irradiation-induced cell damage to the macrophage





# CeONP<sup>60/40</sup> and CeONP<sup>20/80</sup> repress inflammation and osteoclast markers following irradiation-induced cell damage to the macrophage

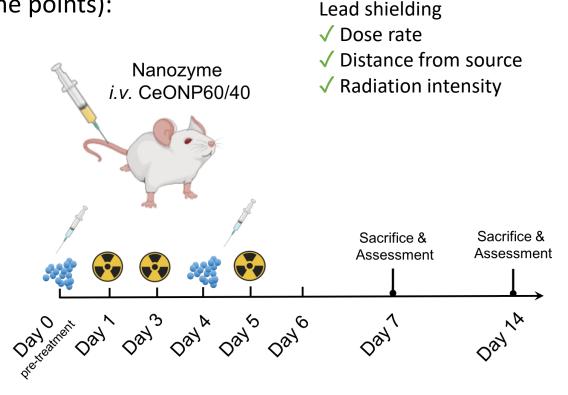




#### *In vivo* studies and analysis of 个Ce<sup>3+</sup>-CeONP in 9-week-old SAS Sprague-Dawley rats and following IR-induced tissue damage

Experimental groups (*n*=6/group; 48 rats in total, two-time points):

- 1) Control
- 2) Control + CeONPs
- 3) X-ray only
- 4) X-ray + CeONPs
- ✓ Histological analysis of kidney, spleen and liver.
- ✓ Complete blood count / blood chemistry.
- ✓ DNA damage (cells in bone marrow niche).
- ✓ Immunohistochemistry (RANKL, senescence).
- ✓ TRAP staining (osteoclastic activity).
- ✓ MicroCT.
- ✓ 3-point bending (fracture stress, ultimate stress).



**Kimtron biological irradiator** 

8 Gy (total 24 Gy)

In rats 7 Gy/day for 5 days (35 Gy) - human equivalent of 70 Gy. Hypofractionated total dose – human equivalent of 48 Gy



### Results: In vivo

✓ Significantly reduced DNA damage to cells within the bone marrow niche.

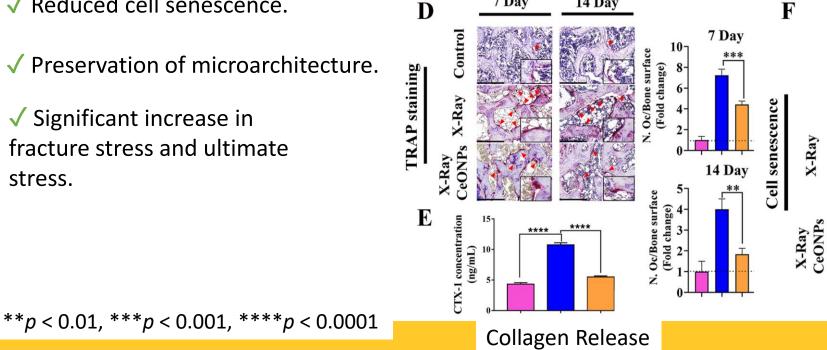
✓ Significantly reduced RANKL from cells within bone.

✓ Significantly reduced osteoclastic activity and CTX-1.

✓ Reduced cell senescence.

 $\checkmark$  Preservation of microarchitecture.

✓ Significant increase in fracture stress and ultimate stress.



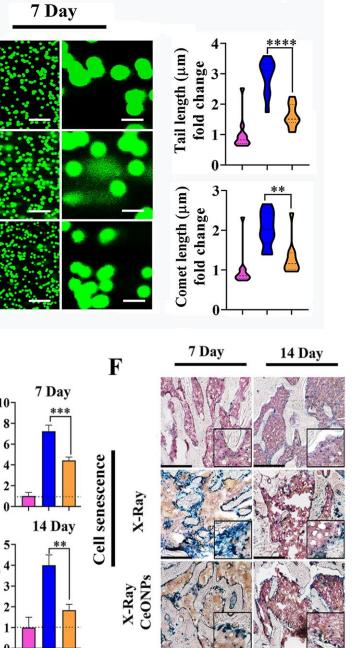
7 Day

Control

X-Ray

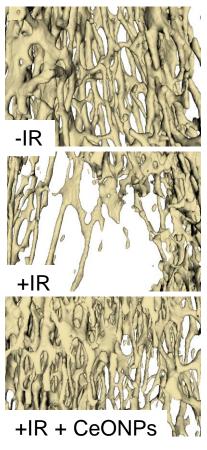
eONPs X-Ray

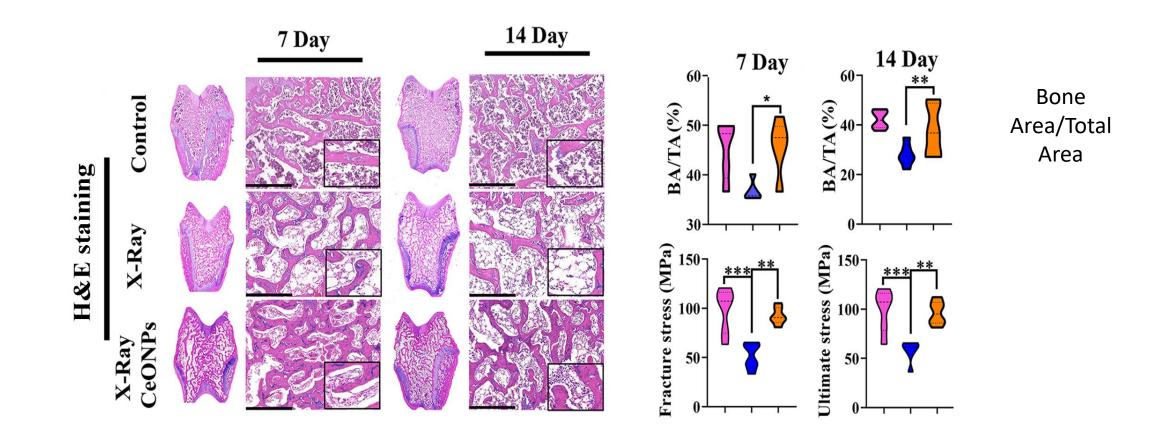
14 Day



#### **Cell Senescence**



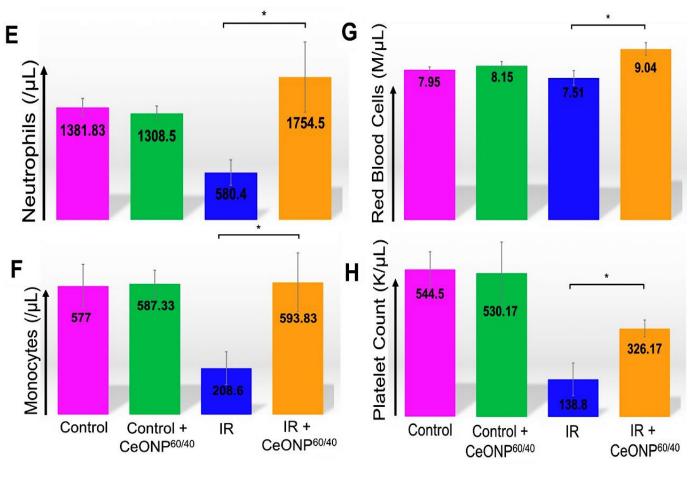




#### CeoNPs maintained bone strength despite exposure to harmful IR.

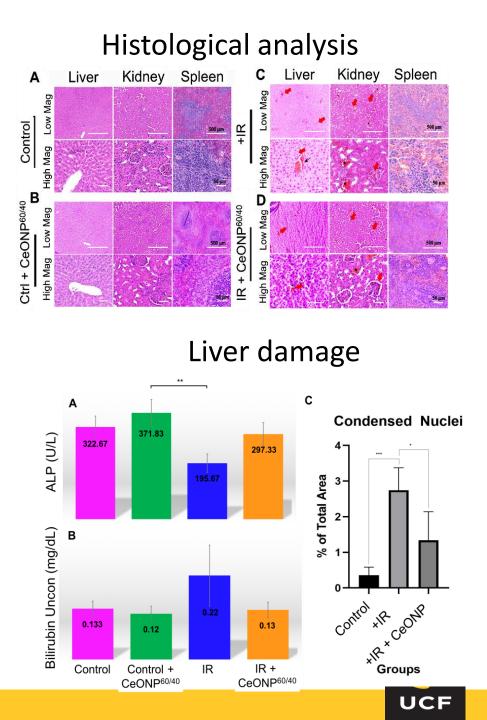


# CeONP<sup>60/40</sup> showed no damage to healthy tissue and protected organs from IR damage



#### Key cells in the blood

p < 0.05, p < 0.01, p < 0.001



# Conclusions

CeONPs were well-tolerated and exhibited a multifunctional protective effect against ionizing radiation-induced damage while augmenting osteogenesis, reducing osteoclastic activity and preventing bone loss
 *Longer term studies are needed*

CeONPs hold promise as a novel multifunctional therapeutic strategy for irradiation-induced bone loss

To the best of our knowledge, this is *first evidence* of CeONPs role in:

✓ Bone regeneration + IR

- $\checkmark$  The critical role of increased Ce<sup>3+</sup> surface sites
- ✓ *Potentially* non-redox related enhanced bone formation









Dr. Fei Wei Dr. Abinaya Pugazhendhi Dr. Mahmoud Omer Christopher Ngo Raven Pascua



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