

Development of Rapid Liquid Scintillation Counting Method for Determination of Tritium in Foods

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Motivations

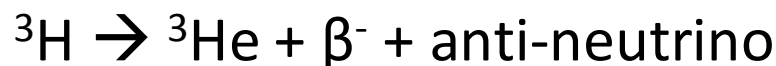
- To improve routine method used for monitoring discharge of ^3H from nuclear power plant into atmospheric, aquatic, and terrestrial ecosystems
- To enable rapid screening of ^3H in foods for prompt decision making in the event of a nuclear or radiological emergency
- To compare performance of different LSC instrument technologies for analyzing ^3H in foods
- To develop a versatile LSC method for detecting ^3H in water and a wide variety of agricultural products
- To determine ^3H baseline radiation by examining ^3H activity in foods purchased from local market

Objectives

- Evaluate and validate distillation procedure to rapidly extract free-water from different types of foods suitable for analysis of ^3H by liquid scintillation counting
- Develop a simple method for rapid and accurate determination of water contents in foods to relate the ^3H activity measured in LSC sample to the ^3H activity presented in foods
- Determine optimum sample and instrument parameters for high throughput ^3H analysis complying with the data quality objectives per regulatory guideline
- Establish a rapid and versatile LSC method suitable for analyzing ^3H in a wide variety of foods and agricultural products

About Tritium (^3H)

^3H decays with a half-life of 12.3 years by emitting a β -particle to form ^3He .



The E_{max} and \bar{E} of β -ray emitted by ^3H are only 18 and 6 keV, which are too low for the radiation to penetrate skin. However, it has the ability to incorporate with the DNA and raise cancer risk when unduly inhaled, ingested, or absorbed over a prolonged period. As a result, its presence in potable water and food products must be monitored per regulatory guidelines.



Current Methods Used for ^3H Analysis

NH State Lab:

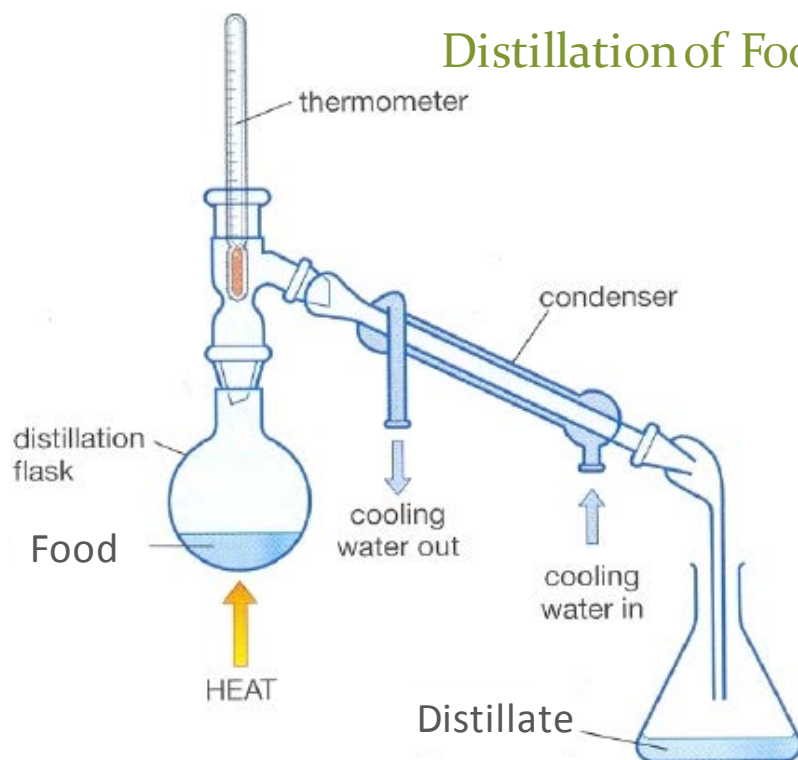
- ✓ Uses heating-mantle distillation method validated for various types of water samples
- ✓ Based on heating sample to release water then collecting condensed vapor
- ✓ Enables quick water extraction from sample
- ✓ Distills water from sample with an open system
- ✓ Accepts large size sample

FDA Rad Lab:

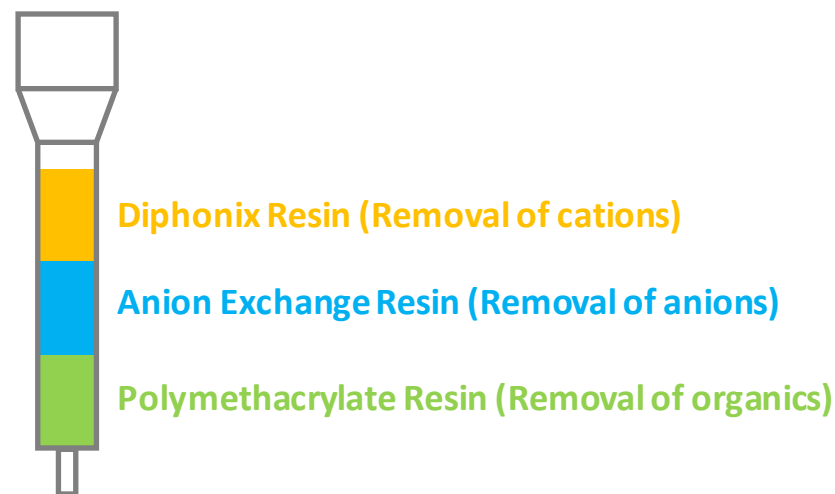
- ✓ Uses vacuum distillation method validated for water and various foods
- ✓ Based on slowly extracting free water from foods
- ✓ Requires over night extraction
- ✓ Extracts water from sample with a closed system
- ✓ Limits sample size to ~30 grams

Extend Heating-Mantle Method for Food Analysis

Free water in foods is readily separable from matrix by heating and reclaiming by condensation. Some color and organic substances may be found in distillate and cause color/chemical quench that biases low measurement results. Treatment of sample distillate with ^3H column may be necessary for certain foods.



Eichrom Tritium Column



Measurement Model

Calculation of ^3H concentration in food:

$$C_T = \frac{R_T \times F_W \times 10}{E_T \times W_{LSC} \times F_Q}$$

where,

C_T = ^3H concentration in food at time of sample analysis, Bq/kg

R_T = Net sample tritium count rate, cps

F_W = Food water content, %

E_T = Tritium counting efficiency, cps/Bq

W_{LSC} = Weight of distillate used for LSC counting, g

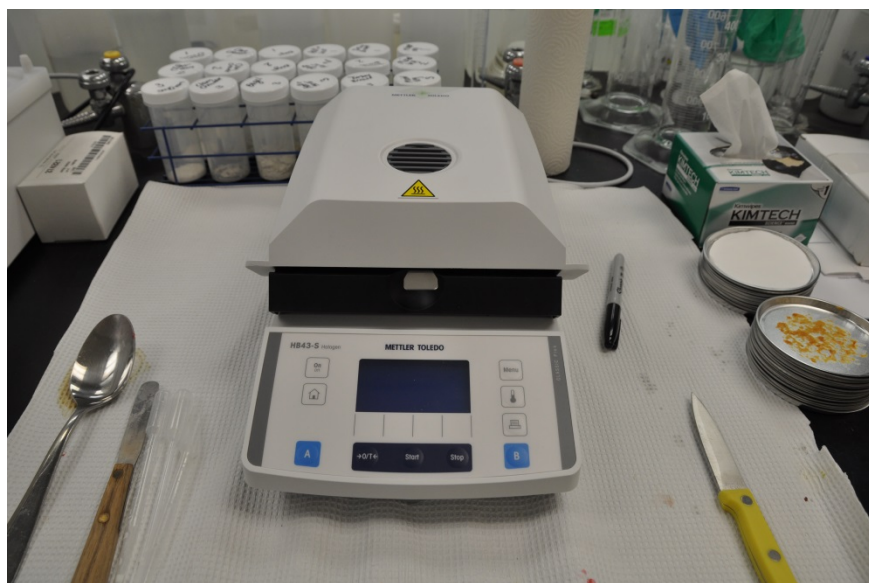
F_Q = Quench correction factor

10 = Conversion factor

Determination of Food Water Content

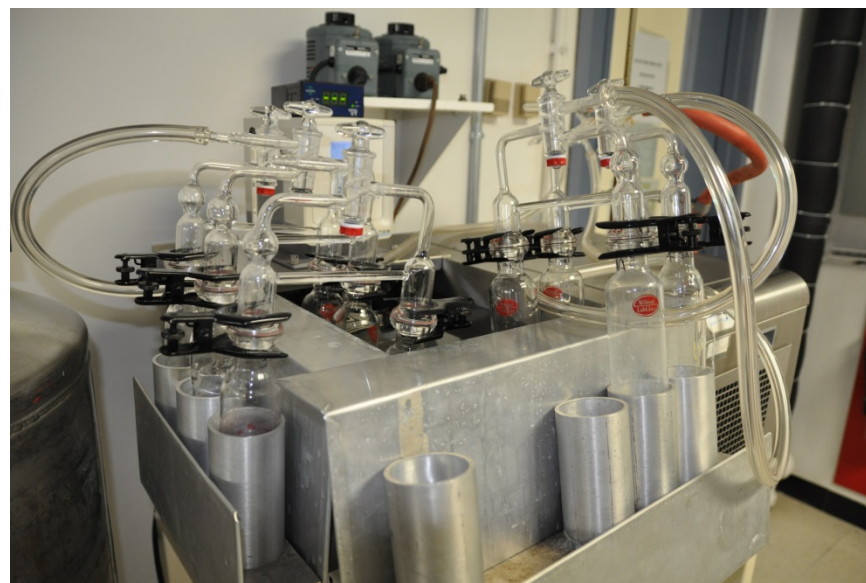
Moisture analyzer was proposed for rapid determination of food water content. To validate its acceptance, the following two methods were used and compared

Moisture analyzer



Quick (~30 min)
Simple

Vacuum distillation



Slow (over night)
Accurate

Comparison of Results





Food water contents
determined by
moisture analyzer and
vacuum distillation

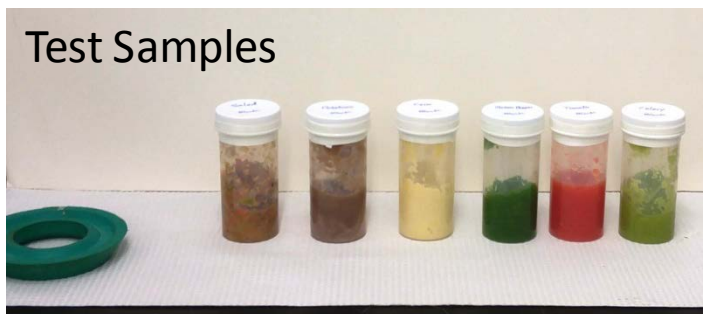
Food	N	Moisture Analyzer	Vacuum Distillation	Diff., %
		Mean \pm 2s		
Lettuces	3	96.35 \pm 0.40	94.95	-1.5
Mushroom	3	91.08 \pm 0.34	89.66	-1.6
Pear	3	83.23 \pm 0.52	83.33	0.1
Cucumber	3	95.66 \pm 0.17	95.32	-0.4
Zucchini	3	94.24 \pm 0.06	93.60	-0.7
Green Beans	2	90.73 \pm 0.18	-	-
Sweet Potato	2	79.00 \pm 0.24	-	-
Greek Yogurt	2	76.78 \pm 1.78	-	-
Watermelon	2	91.61 \pm 0.04	-	-
Whole Milk	2	88.04 \pm 0.13	-	-
Mustard Green	3	89.25 \pm 0.34	87.01	-2.6
Chard	3	91.13 \pm 0.51	92.80	1.8
Grape	3	82.58 \pm 0.52	82.06	-0.6
Orange	3	86.06 \pm 0.34	84.13	-2.3
Fish	3	77.64 \pm 0.51	77.25	-0.5
Ground Beef	3	60.11 \pm 2.02	61.46	2.2
Ground Pork	3	62.62 \pm 1.47	64.77	3.3
Strawberry	3	89.14 \pm 0.29	88.78	-0.4
Corn	3	69.75 \pm 0.12	69.16	-0.8
2% Milk	3	89.15 \pm 0.11	87.77	-1.6

Test Materials and Method Procedure

List of matrix blanks and matrix spikes used

Food	Weight, g	Food	Weight, g
Corn	65.21	Strawberry	65.55
Celery	65.47	Blueberry	65.69
Potatoes	65.27	Green Beans	65.64
Tomato	65.45	Sweet Potato	67.66
Green Pepper	65.18	Fish	60.64
Mixed salad	65.96	Apple	70.40
		Plain Greek Yogurt	66.85
		Watermelon	66.37
		Whole Milk	66.30
		2% Milk	65.26
		Corn	69.28

Preparation of sample for ^3H analysis



Each test sample was used entirely



Distillation of foods were found to be mostly straightforward but certain types of foods need additional treatment to obtain clear and colorless distillate before LSC counting.

Distillation System Setup

- Capable of distilling 9 samples at a time within one hour
- Assembled inside a fume hood to prevent ^3H from escaping into the room



Materials and Method (Cont.)

- Figure of Merit:

$$FOM = \frac{(Efficiency^2) \cdot (Volume^2)}{Background}$$

- 8 mL sample + 12 ml of Ultima Gold LLT
- Teflon-coated plastic vial
- Efficiency = 43% (Hidex 300SL)
- Measurements done with and without dark adaptation: the activities for non-dark adapted blank samples were ~10% higher compared to the 2-hour dark adapted ones

Removal of Sample Color/Cloudiness

- Eichrom's Tritium column was used to treat colored and/or cloudy sample distillate to eliminate the need for repeating the distillation, thus expedite the sample preparation



Potato

Before treatment



Potato

After treatment

Comparison of Instruments

- Liquid scintillation counters used:

- Hidex 300SL
- Quantulus 1220
- TriCarb 3170 TR/SL



Hidex 300SL

- Count time:

- 100 min – Hidex
- 60 min – Quantulus 1220
- 60 min – TriCarb 3170 TR/SL



TriCarb 3170 TR/SL

- Background suppression:

- TDCR + Digital shielding + Cooling – Hidex
- Anti – coincidence + Pb Shield – Quantulus 1220
- Anti – coincidence + BGO Shield – TriCarb 3170 TR/SL



PerkinElmer

Quantulus 1220

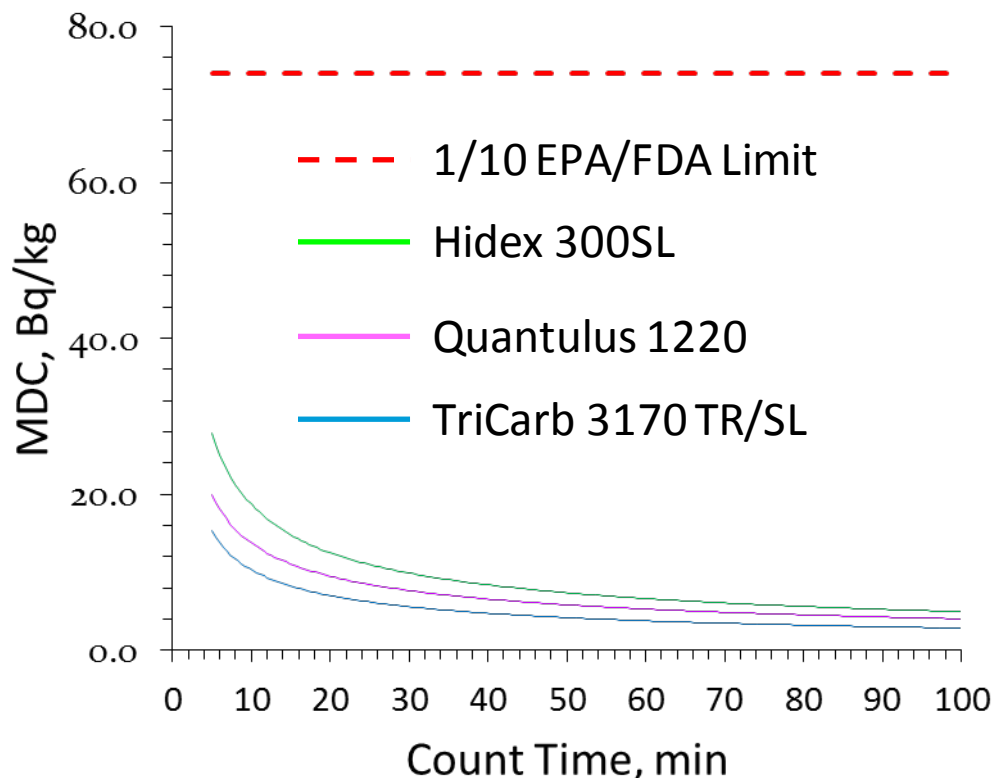
Comparison of ^3H Matrix Blank Count Rates

Food Matrix	Hidex 300SL, cpm	Quantulus 1220, cpm	TriCarb 3170 TR/SL, cpm
Celery	6.95 ± 0.68	3.20 ± 0.47	0.72 ± 0.41
Potatoes	7.45 ± 0.70	3.03 ± 0.45	0.15 ± 0.37
Tomato	7.02 ± 0.68	3.20 ± 0.47	0.56 ± 0.40
Green Pepper	7.19 ± 0.69	3.30 ± 0.47	0.41 ± 0.39
Mixed Salad	7.07 ± 0.69	3.01 ± 0.45	0.51 ± 0.39
Watermelon	6.58 ± 0.66	3.33 ± 0.48	0.60 ± 0.41
Blueberry	6.05 ± 0.64	3.15 ± 0.46	0.52 ± 0.41
Strawberry	7.05 ± 0.68	3.90 ± 0.52	0.49 ± 0.41
2% Milk	7.73 ± 0.72	3.71 ± 0.50	0.67 ± 0.42
Whole Milk	6.80 ± 0.67	3.25 ± 0.47	0.31 ± 0.39
Corn	6.89 ± 0.67	4.02 ± 0.52	0.15 ± 0.39
Fish	7.33 ± 0.70	3.49 ± 0.49	0.32 ± 0.38
Sweet Potato	7.45 ± 0.70	3.32 ± 0.48	0.25 ± 0.39

All uncertainties are given at 95% confidence level

Estimated Detection Limits vs EPA/FDA Limit

- Sample volume = 8 mL
- Ultima Gold LLT = 12 mL
- Method blank = ~7 cpm Hidex 300SL (Eff = ~43%)
- Method blank = ~2 cpm Quantulus 1220 (Eff = ~29%)
- Method blank = ~0.5 cpm TriCarb 3170 TR/SL (Eff = 21% UltraLow level mode)



Observed Sample Quench Effect

Food Matrix	Quantulus 1220 Quench Index, SQP	TriCarb 3170 TR/SL Quench Index, tSIE
Celery	715.94	349.52
Potatoes	714.50	345.89
Tomato	722.49	368.72
Green Pepper	718.76	360.54
Mixed Salad	723.60	371.24
Watermelon	724.75	375.57
Blueberry	725.44	384.88
Strawberry	723.17	379.12
2% Milk	723.77	376.89
Whole Milk	727.39	384.67
Corn	701.13	311.20
Fish	725.45	383.03
Sweet Potato	698.34	313.27

Abnormal sample quench and instability were observed for foods with high starch contents, which biased the results without polishing the sample distillate with Eichrom tritium column

Comparison of ^3H Results

Food Matrix	Known Bq/kg	Hidex Bq/kg	Quantulus 1220 Bq/kg	TricCarb 3170 Bq/kg
Celery	636.18 \pm 7.84	603.07 \pm 11.49	651.30 \pm 17.45	604.96 \pm 19.9
Potatoes	638.13 \pm 7.87	622.20 \pm 10.69	469.24 \pm 13.42	468.04 \pm 16.0
Tomato	636.37 \pm 7.84	651.07 \pm 11.86	679.39 \pm 17.66	651.48 \pm 20.5
Green Pepper	639.01 \pm 7.88	657.06 \pm 11.79	643.52 \pm 16.99	634.18 \pm 20.0
Mixed Salad	631.45 \pm 7.78	629.69 \pm 11.79	676.57 \pm 17.86	658.44 \pm 20.9
Watermelon	502.04 \pm 6.19	457.24 \pm 9.49	520.18 \pm 15.29	501.52 \pm 17.55
Blueberry	253.62 \pm 3.13	274.85 \pm 7.29	272.22 \pm 10.45	264.12 \pm 12.36
Strawberry	254.16 \pm 3.13	260.63 \pm 7.20	278.80 \pm 10.71	258.52 \pm 12.41
2% Milk	638.22 \pm 7.87	619.17 \pm 10.84	707.36 \pm 17.83	632.82 \pm 19.49
Whole Milk	628.21 \pm 7.74	626.41 \pm 10.79	681.71 \pm 17.18	621.47 \pm 19.08
Corn	601.19 \pm 7.41	563.68 \pm 9.73	479.82 \pm 13.63	439.65 \pm 15.26
Fish	549.48 \pm 6.77	634.32 \pm 10.37	565.13 \pm 14.92	527.35 \pm 16.77
Sweet Potato	246.23 \pm 3.04	212.43 \pm 6.49	211.96 \pm 8.82	194.76 \pm 10.30

All uncertainties are given at 95% confidence level

Conclusions

- A moisture analyzer is reliable and accurate for determination of food water content needed to relate the ^3H activity measured in LSC sample to the ^3H activity presented in foods
- The preliminary results were found to be acceptable per FDA's data quality objective
- Considering the sensitivity, rapidness, and simplicity achieved, the method is suitable for high throughput ^3H analysis
- The method can eliminate color quench and maintain constant sample quench using Eichrom tritium column when necessary

Conclusions (Cont.)

- Additional study is needed to fully validate the method with a wider range of food matrices
- Up to 36 food samples can be prepared for LCS counting per day
- Despite that the method presented an alternative approach for analyzing ^3H in foods, additional studies on the method performance characteristics are still needed before official use

Acknowledgement

We would like to thank Lablogic Systems, Inc. (Hidex) for assisting instrument optimization and use of application software during the study.

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Thank you!

Any questions?



